

Scale dependency of processes structuring metacommunities of cladocerans in temporary pools of High-Andes wetlands

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Metacommunity structure can be shaped by a variety of processes operating at different spatial scales. With increasing scale, the compositional variation among local communities (beta diversity) may reflect stronger environmental heterogeneity, but may also reflect reduced exchange of organisms between habitat patches. We analyzed the spatial architecture of a metacommunity of cladoceran zooplankton in temporary pools of High Andes wetlands, with the objective of explaining the spatial dependency of its structure. The spatial distribution of the pools is hierarchical and highly discontinuous: pools are clustered within small wetlands, which lay scattered over valleys that are separated from each other by mountain ridges. We studied a total of 59 pools, belonging to six different wetlands in four different valleys. We assessed pool environmental heterogeneity and sampled active communities and dormant propagule banks of cladoceran zooplankton. Environmental heterogeneity proved very high within wetlands and showed almost no increase with increasing spatial scale. Conversely, diversity partitioning analyses indicated an increase in beta diversity with spatial scale, especially among valleys. Variation partitioning on environmental data and spatial RDA models suggested environmental heterogeneity as the most important generator of beta diversity within wetlands. At the largest spatial scale, beta diversity manifested itself mainly as a differentiation of species occurrence patterns among valleys, which could not be entirely explained by environmental variables. Our study thus presents a case where environmental control seems to be the dominant metacommunity structuring process at the smallest spatial scale, whereas neutral processes and dispersal limitation are the most likely generators of beta diversity at the largest spatial scale.

The importance of spatial scale has increasingly been recognized in community ecology, especially during the latest decades (Menge and Olson 1990). Compositional variation among communities (beta diversity) can be generated by different factors and the relative importance of these factors can vary among spatial scales. One important reason for this is that environmental variables that shape communities may differ in their range of variation among spatial scales. Some variables show large variation at small spatial scales and will therefore generate high community dissimilarity in relatively small areas. Other important variables only show substantial variation at large spatial scales, and will thus only give rise to community variation in large study areas (Borcard et al. 2004).

A second major driver of beta diversity is dispersal limitation. Since the breakthrough of the metacommunity concept (Leibold et al. 2004, Holyoak et al. 2005), community ecology has increasingly considered alternative models of spatial dynamics in explaining patterns of diversity within and among communities at the landscape scale. Dispersal plays a pivotal albeit different role in all these models. Neutral and patch dynamics models, which assume absence of species interactions with the environment (Hubbell 2001), strongly rely on dispersal limitation to explain patterns of (meta-) community structure. Conversely, the species sorting model (sensu Leibold et al. 2004, Cottenie 2005, also referred to here as "environmental control") explains community composition by the interaction of species niches with the abiotic and biotic environment and assumes that dispersal limitation does not prevent species from tracking environmental gradients in space and time. In the case of mass effects, massive fluxes of individuals from source to sink communities can overwhelm the effect of local conditions and species interactions (Vanschoenwinkel et al. 2007, Guelat et al. 2008). The potential of organisms to disperse among habitat patches within metacommunities depends on the distance and type of connections among patches (Shurin et al. 2009). As spatial scale is intrinsically related to the among-patch distances and the physical structure of the landscape, it is expected to be very important in determining the type and

strength of alternative types of spatial dynamics that shape metacommunity structure (Dumbrell et al. 2008).

The revival of diversity partitioning techniques (Lande 1996, Veech et al. 2002, Jost 2007) has strongly enhanced research on the spatial organization of biodiversity in metacommunities. For spatially nested datasets, these techniques allow researchers to decompose overall gamma diversity (total regional diversity) into alpha diversity (mean local diversity) and components of beta diversity for each level of spatial scale. Although this approach has considerably increased our knowledge on the overall architecture of metacommunity biodiversity for several organism groups in a variety of landscape contexts (Stendera and Johnson 2005, Diekotter et al. 2008, Lindo and Winchester 2008), it often has limited power in explaining the observed patterns when standing on its own. Parallel to the development of diversity partitioning, an increasing number of studies have tried to link patterns of community variation with existing metacommunity paradigms (Holyoak et al. 2005) by partitioning community variation into spatial and environmental components (Borcard et al. 1992, Cottenie 2005), using direct gradient ordination techniques (Legendre et al. 2005). These studies have increased our understanding of processes underlying metacommunity patterns, but the large majority of these studies are confined to a single spatial scale. In order to get a better understanding of the link between the structure and dynamics of metacommunities at different levels of spatial scale, there is a need for an integrated approach through the combined application of these two analytical frameworks.

Due to their discontinuous distribution at multiple spatial scales, temporary pools of small wetland systems in the high Andes provide an interesting model system for the study of scale dependency in aquatic invertebrate metacommunities. The spatial distribution of pools in this part of the Andes is hierarchical: pools are clustered within small wetlands and are to a variable degree connected with each other during periods of high rainfall. These wetlands lay scattered over valleys that are separated from each other by mountain ridges with height of ca 450 m. For this study, we focused on cladoceran zooplankton, a group of passively dispersing aquatic invertebrates (Bohonak and Jenkins 2003) for which time-integrated species lists of individual pools can readily be obtained through the analysis of dormant propagule banks (Vandekerkhove et al. 2005a, b). We sampled pools at three levels of spatial scale (i.e. withinwetlands, within valleys and among valleys) with the aim of dissecting the spatial architecture of the metacommunity and evaluating the relative importance of alternative metacommunity processes for each level of spatial scale. We expected beta diversity among pool communities to increase with increasing spatial scale and set out to explore if that increase corresponded to community patterns that should emerge from stronger environmental control along more pronounced environmental gradients, or whether these patterns were more consistent with the patterns that were expected to emerge as a result of stronger dispersal limitation. At the smallest spatial scale, the scale of individual wetlands, we expect little dispersal limitation, given the small size of the wetlands and short distances among pools; conversely, flooding events may lead to homogenization of communities through mass effects. At

the intermediate and largest spatial scales, we expect an increase in the importance of dispersal limitation, given the larger among-patch distances, lower hydrological connectivity and the presence of mountain ridges between valleys. In addition, broader environmental gradients may potentially enhance beta diversity at these larger scales, as long as environmental control is not impeded by dispersal limitation (Leibold and Norberg 2004).

Material and methods

Study area

Our study area is located in the Tunari mountain range (Cordillera del Tunari, between 17°19'19'' and 17°10' 56''S and 66°08'53" and 66°22'43"W), at altitudes between 4000 and 4400 m, and is part of the eastern Andes mountain range (Fig. 1). It is a mountainous area, with numerous small wetlands (locally called "bofedales") that lay scattered over the valleys and mountain slopes (Coronel et al. 2004). Most of the wetlands in this area contain small temporary fishless pools, of which the total number typically varies between one and eight, although pools can be more numerous in some of the larger wetlands. The region is subject to a rainy season from October to March and a dry season from April to September. During the latter period, the pools fall dry. We refer to Coronel et al. (2004) for a more detailed description of the limnological features of the pools.

Sampling design

We sampled a total of 59 pools in four different valleys of the Tunari mountain range, i.e. Taquiña, Toro, Saito and San Ignacio. The aim of our sampling design was to study cladoceran community variation at three levels of spatial scale, i.e. at the level of individual wetlands, individual valleys and multiple valleys. Full hierarchical sampling would have required us to sample all pools in the entire set of wetlands in each of the studied valleys, an effort that was unfeasible in the context of the present study. We therefore applied a different approach still allowing us to capture variation among communities at the three levels of spatial scale. For this, we collected two sets of samples (Fig. 1). For a first dataset (further referred to as the WTL dataset), we sampled all pools in 6 different wetlands, i.e. two wetlands in Taquiña (Tq1, with 6 pools and Tq2, with 7 pools), two wetlands in Toro (Tr1, with 6 pools and Tr2, with 5 pools), one wetland in Saito (S1, with 5 pools) and one wetland in San Ignacio (SI1, with 6 pools). The total number of pools sampled for the WTL dataset thus equals 35 from 6 wetlands (Fig. 1). For the second dataset (the VALLEY dataset), we selected a number of wetlands in each of the four valleys (13 wetlands in Taquina, 7 in Toro, 5 in Saito and 4 in San Ignacio) and in each of these wetlands we sampled one haphazardly chosen pool (Fig. 1), including one randomly selected pool of each wetland from the WTL dataset. In total, the VALLEY dataset consists of 29 sampled pools from 29 wetlands. The WTL dataset thus represents cladoceran community variation within wetlands, whereas the VALLEY dataset represents community



Figure 1. Location of the four valleys sampled in the Tunari mountain range (Cochabamba, Bolivia) and a schematic representation of the wetlands (represented by ovals) and pools (represented by circles) sampled in each valley. Filled symbols represent pools sampled for the VALLEY-dataset. Empty circles represent pools that were additionally sampled for the WTL-dataset.

variation among wetlands at the valley scale. When combined, the WTL and VALLEY dataset also represent variation among valleys at the regional scale; there are 59 pools from 29 wetlands located in 4 valleys.

Sampling

We assessed a variety of pool characteristics during a sampling campaign in February–March 2004, in the middle of the wet season. We determined the concentration of nitrates (NO₃), total phosphate (TP), alkalinity, pH, conductivity, macrophyte coverage, phytoplankton chlorophyll a, thickness of the sludge layer on the sediments, pool depth and pool surface area. In addition, we assessed the density of potential predators (cyclopoid copepods, mites, and larvae of the coleopteran genera *Ranthus* (Colymbetinae) and *Hydroporus* (Hydroporinae). We refer to Coronel et al. (2004) for methodological details on the collection of the data for these variables.

We collected cladoceran community data in two different ways: 1) by taking snap shot samples of active communities at one sampling occasion, and 2) through the analysis of dormant egg banks. These two methods have been shown to be complementary in their ability to detect cladoceran species (Vandekerkhove et al. 2005a). We sampled active communities during February–March 2004 by collecting water with a tube sampler (75 mm diameter and 1.5 m length) from different places (vegetated and non-vegetated areas) in the pool and by filtering this water through a 30- μ m Nitex mesh. The total filtered volume ranged between 3 and 15 L. Samples were preserved in sucrose-formaldehyde solution (5% final concentration). We collected sediments with dormant eggs at the beginning of the dry season of 2006 (26–29 June). In each pool, we collected sediment at ten haphazardly chosen locations using a KC-sediment core sampler (0.7 m long plexi-glass tube of 5.2 cm diameter). Only the upper three centimeters of each core were retained (ca 100 g wet weight per sample). A total amount of one kilogram sediment per pool was collected. Immediately after collection, samples were wrapped in aluminum foil and transported to the lab in a cooler box.

Sample analysis

We analyzed active community samples by counting a total of at least 300 individuals from sub-samples. For the analysis of the sediment samples, we first isolated dormant eggs by means of the Onbé-Marcus method as modified by Vandekerkhove et al. (2004). After storage in the dark at 4°C, we first removed gross material (mostly vegetal debris) using sieves of 1 mm and 500 μ m. Dormant eggs retained by a sieve of 63- μ m mesh were then isolated from fine debris by the sugar flotation method, following

three steps: 1) filtration through a 48-µm mesh, 2) centrifugation of the residual in a sugar solution (1000 g table sugar in 1000 ml distilled water) at 3000 rpm for three minutes, and 3) washing of the supernatant over a 48-µm size mesh using tap water. The isolated dormant eggs (at least 300 eggs whenever possible) were next sorted according to morphology, identified and counted under a stereo microscope (Olympus SZX12). To allow identification to species level, we incubated unknown dormant egg types individually in 30-ml multi-well plates containing the Aachener Daphnien Medium (ADAM medium) at a conductivity of 30 μ s cm⁻¹. Multi-well plates were placed in an incubator at 20°C with a photoperiod of 14 h light and 10 h dark. Incubation medium was refreshed every five days. For a period of two months, we checked all multiwell plates every four days for emerging hatchlings. Hatchlings were transferred to 50-ml vessels and fed *Scenedesmus obliquus* (100 000 cells ml⁻¹) until a developmental stage at which they could be identified. We used the keys of Pagui (1995) and Smirnov (1996) for identification.

Data analysis

We compiled species lists for each of the pools, combining data from the active community snapshot samples and the dormant propagule banks, and derived a presence-absence dataset from these lists. We also calculated relative abundances of species using the data from the snapshot samples of active communities.

Geographic distance and environmental heterogeneity across spatial scales

We calculated the mean geographic distance among pairs of pools for each of the three levels of spatial scale, i.e. among pools within each individual wetland (WTL dataset), among pools belonging to different wetlands within each valley (VALLEY dataset) and among pools belonging to different valleys (WTL and VALLEY datasets combined). In the same way, we used the environmental variables to estimate environmental heterogeneity among pairs of pools at the within-wetland, the within-valley and the amongvalley scales as standardized Euclidean distances.

Beta diversity of cladoceran communities across spatial scales

We estimated beta diversity at each spatial scale following two different approaches: 1) as the mean Bray-Curtis dissimilarity among pairs of pools, similar as for geographic distance and environmental heterogeneity (i.e. among pools within wetlands using the WTL dataset; among pools from different wetlands within valleys using the VALLEY dataset, and among pools from different valleys). These calculations were done on the presence-absence data derived from the species lists as well as on the relative abundance data of the active communities; 2) through the application of diversity partitioning on species richness of the species lists and true Shannon diversity (Jost 2007) estimated from the abundance data of active communities. Species richness was partitioned in the additive way, with $\gamma = \alpha + \beta 1 + \beta 2 + \beta 3$ (Lande 1996), where α refers to the average richness in local communities (samples), gamma refers to the total richness observed in the entire set of samples, and ß1, ß2 and ß3 refer to the beta diversity at each of the studied spatial scales: $\beta_1 = among$ pools within wetlands; $\beta_2 = among$ wetlands within valleys; f3 = among valleys. Additive diversity partitioning has the advantage that beta components can directly be quantitatively compared with each other and with alpha and gamma diversity. True Shannon diversity can only be partitioned in a multiplicative way, but we transformed its alpha, beta and gamma components logarithmically (i.e. into Shannon entropies) so as to make them relate additively (Jost 2007). A full hierarchical partitioning analysis, including the three levels of spatial scale, could only be performed on a subset of the WTL dataset, including all data of Taquina and Toro, but excluding the data of Saito and San Ignacio. Incorporation of the latter valleys in the analysis would have resulted in a bias, underestimating average beta diversity among wetlands (ß2), because there were only data available for one wetland in each of these valleys. To assess the robustness of these results and fully exploit the size of our dataset, we performed additional partitioning analyses on our complete dataset, estimating beta diversity among pools within each of the wetlands in the WTL dataset (ß1) as well as among pools belonging to different wetlands in each of the valleys of the VALLEY dataset. Logically, the latter variable incorporates both within- and among-wetland beta diversity within valleys and should equal \$1 + \$2. Diversity partitioning calculations were performed with the software PARTITION v3 (Veech and Crist 2009).

Explaining the beta components

A commonly applied method to quantify the relative importance of alternative metacommunity processes consists of the decomposition of community variation into an environmental (E) and a spatial (S) component. Using direct gradient ordination techniques (mostly redundancy analysis), statistical models are first constructed for both components and their relative contributions to community variation are subsequently assessed through variation partitioning (Borcard et al. 1992). Upon correction for the spatial component, a significant environmental component (E/S) is generally accepted to represent environmental control. A significant spatial component, after control for the environmental component (S/E), can indicate neutral processes, historic events and dispersal limitation (Legendre and Legendre 1998, Cottenie 2005), assuming that all important environmental variables have been measured. The co-occurrence of significant environmental and spatial components has been considered as being indicative of mass effects (Cottenie 2005), although it can also represent species sorting that is partially constrained by dispersal limitation (Ng et al. 2009).

To explain beta diversity in the cladoceran communities, we applied variation partitioning on redundancy analysis models (RDA) for each of the three levels of spatial scale, separately. For the largest spatial scale, i.e. the inter-valley scale, we constructed an environmental and a spatial model using the entire dataset (VALLEY and WTL datasets combined, n = 59). The environmental model (E) was constructed by applying the forward selection procedure of Blanchet et al. (2008) to the environmental variables. For the spatial model (V), we constructed a matrix representing valley identity using 3 dummy variables. With variation partitioning (Borcard et al. 1992, Legendre et al. 2005, Peres-Neto et al. 2006), we then tested the marginal and unique contributions of the environmental and spatial models in explaining community variation.

We applied the same procedures to the VALLEY dataset (n = 29) for the study of beta diversity at the withinvalley scale. However, the spatial model consisted here of two components: 1) a V-component, representing valley identity using dummy variables (see above), and 2) an S-component consisting of Moran's eigenvector maps (MEM) that describe the spatial relationships among pools within individual valleys. MEM analysis produces a set of orthogonal spatial variables, derived from the geographic coordinates of the study sites (Dray et al. 2006). These variables represent spatial variation across a range of spatial scales and can be used as explanatory variables in direct gradient analysis to model spatial relationships in community data. In the S-matrix, the MEM variables were arranged in blocks, each block corresponding to one valley. Within these blocks, pools from the other valleys received the mean value 0. The structure of the blocks of MEM variables in S were similar to that used to test the presence of different spatial structures at different times, shown in Appendix C of Legendre et al. (2010). Supplementary material Appendix 1 provides an R function that can be used for the construction of a staggered matrix of MEM spatial eigenvectors for sites that are spatially clustered. The type of MEM variables computed in the present study were formerly called principal coordinates of neighbour matrices (PCNM: Borcard and Legendre 2002, Borcard et al. 2004, Dray et al. 2006).

Using the VALLEY dataset, we then applied variation partitioning to assess the unique contributions of the Sand E- model components to community variation, while controlling for the V-component. An identical procedure was applied for studying cladoceran variation at the intrawetland scale using the WTL dataset (n = 36).

All analyses were carried out on the species lists (active and dormant community data combined) as well as on the relative abundance data from the active communities. We did the analyses on these two datasets because they should cast light on different aspects of metacommunity dynamics. Indeed, we expect that analyses on abundance data of active communities should mainly stress the impact of environmental gradients or mass effects, because they emphasize shifts in the relative success of species and also because these data were collected simultaneously with the environmental data. Analyses on species lists should do better at reflecting distribution patterns of species; we expect them to be more powerful in revealing patterns related to dispersal limitation.

All species data (both abundance and presence-absence) were Hellinger transformed prior to analysis (Legendre and Gallagher 2001). After this transformation, RDA and variation partitioning are based on the Hellinger distance, which is appropriate for community composition data (presence-absence or abundance), instead of being based on the inappropriate Euclidean distance; the Hellinger distance computed on presence-absence data is monotonically related to the Ochiai distance, which is also appropriate for community composition data. Of the environmental variables, conductivity, surface area, sediment depth and chlorophyll a were log-transformed. Densities of predatory invertebrate taxa tended to be positively correlated with each other; we summarized overall predator density with the first principal component (i.e. sample scores vector) of a standardized principal component analysis (eigenvalue: 39%). All statistical analyses were performed in R (v2.8.1; R Development Core Team 2008), using the rda and varpart functions of the vegan package (Oksanen et al. 2005), the forward.sel function of the packfor package (Dray et al. 2007); MEM spatial eigenfunctions were computed using the PCNM function of the PCNM package (Legendre et al. 2009).

Results

Geographic distance and environmental heterogeneity across spatial scales

The mean geographic distance among pools increased by more than one order of magnitude between successive levels of spatial scale (Fig. 2A). The distance among pools within wetlands (cf. WTL dataset) averaged 0.063 km. The mean distance among pools of different wetlands within valleys (cf. VALLEY dataset) was 0.69 km, whereas pools from different valleys were located at an average distance of 19.7 km from each other (WTL and VALLEY datasets combined).

The mean Euclidean distances for environmental variables were highly variable, both among individual wetlands as well as among valleys (Fig. 2B). We detected no tendency for higher heterogeneity at the valley scale compared to the wetland scale. Heterogeneity among pools from different valleys also tended to be only slightly higher than the within-wetland or within-valley means. See Supplementary material Appendix 2 for summary statistics of the environmental variables.

Patterns of beta diversity across spatial scales

Although highly variable among wetlands and among valleys, mean Bray-Curtis dissimilarities based on species lists tended to increase with spatial scale (Fig. 2C). In contrast, no such tendency was observed for Bray-Curtis dissimilarity based on the relative abundance of species in active communities (Fig. 2D).

A full hierarchical diversity partitioning analysis on the richness data of the Taquina and Torro subset showed a substantial contribution of both the smallest ($\beta_1 = 7.2$) and largest spatial scales ($\beta_3 = 5.5$ species) to gamma diversity (Fig. 3A, left pane). Beta diversity among wetlands within valleys ($\beta_2 = 3.5$ species) was relatively small. The additional partitioning analyses on the WTL and VALLEY datasets (Fig. 3A, right pane) also showed a relatively high average beta diversity among pools within wetlands (β_1)



Figure 2. Average geographic distance (A), environmental heterogeneity (B) and community dissimilarity (C, D) among pools across the three levels of spatial scale (i.e. within wetlands, among pools of different wetlands within valleys and among pools of different valleys). Environmental heterogeneity was calculated as the standardized Euclidean distance for the environmental variables. Community dissimilarities were calculated as the Bray-Curtis distance among pools based on species lists (presence-absence data derived from the combined data of active communities and dormant propagule banks, C) as well as on the relative abundance data of the active communities (D). Stars represent individual wetlands and valleys; circular symbols represent averages and error bars denote the standard deviation.

compared to the average beta diversity among pools from different wetlands at the valley level ($\beta_1 + \beta_2$). According to a paired t-test, both estimates of beta diversity could not be shown to differ significantly, indicating only a minor contribution of β_2 to gamma diversity.

We observed similar patterns for Shannon diversity of active communities (Fig. 3B). Full hierarchical diversity partitioning also indicated a small contribution of β_2 and a relatively large contribution of β_1 and β_3 . Average beta diversity at the valley level ($\beta_1 + \beta_2$; cf. VALLEY dataset) showed no significant difference with the average beta diversity within wetlands (β_1 ; WTL dataset) according to a paired t-test (p > 0.05).

Community variation and environmental gradients

Supplementary material Appendix 3 gives a list of the species and their abundances in samples of active communities and dormant propagule banks. Species lists differed significantly among valleys (V- and V/E-components in

Table 1A), with valley identity explaining up to 10.6% of the community variation. pH proved to be the only significant environmental variable (cf. E in Table 1A), although the effect of this variable was strongly reduced upon correction for inter-valley differences (E/V in Table 1A left). Within valleys and wetlands, MEM models showed no evidence for spatial patterns. Environmental models explained significant fractions of the variation in the species lists of the WTL and VALLEY datasets, but these effects became non-significant when the identity and MEM models of valleys and wetlands were taken into account, respectively (Table 1B and C left).

At the inter-valley scale, the composition (in terms of relative abundance) of the active cladoceran communities was significantly explained by both the environmental model (consisting of the variables alkalinity, sludge depth and surface area) and by valley identity, although the environmental model explained approximately 3 times more variation than valley identity (see E-, V-, E/V- and V/E-components in Table 1A right). Analyses at the withinvalley level using the VALLEY dataset (Table 1B right)



Figure 3. Diversity partitioning results for species richness (A) and Shannon entropy (B). The left panels in the graphs represent the results of a full hierarchical analysis on the Taquina and Toro subset. The right panels represent average results for each wetland and each valley in the WTL- and Valley-datasets, respectively. Alpha: local pool diversity; Beta1: beta diversity among pools within wetlands; Beta2: beta diversity among wetlands within valleys; Beta3: beta diversity among valleys. Beta diversity among pools in the VALLEY-dataset comprises both Beta1 and Beta2 and is therefore denoted as Beta1+Beta2. Error bars represent the standard deviation.

revealed no significant spatial patterns, whereas the explanatory power of the environmental model was low (only marginal effects were significant). At the within-wetland level (WTL dataset, Table 1C right), the marginal and conditional effects of the environmental model (based on alkalinity, surface area, sludge depth and the predation gradient) amounted to 21 and 17% of explained variation, respectively. At this level of spatial scale, the MEM model explained no community variation.

Discussion

In our study, geographic distances among pools increased with more than one order of magnitude across each level of spatial scale. This, however, translated only in a weak increase in environmental heterogeneity and some individual wetlands encompassed a degree of pool heterogeneity similar to what was present at the landscape scale. We found this high within-wetland heterogeneity to concur with relatively high cladoceran beta diversity, both in terms of species richness and Shannon diversity. With RDA analysis, we were able to uniquely explain part of the variation among active communities by environmental gradients, which suggest that this beta diversity at the wetland scale is structured by environmental heterogeneity (Cottenie and De Meester 2004). Part of the unexplained beta diversity may also have originated historically by chance (cf. priority effects; Louette and De Meester 2007, Loeuille and Leibold 2008), although we have no specific data to further support this idea. We found no evidence for spatial community patterns within wetlands.

Beta diversity at the valley scale was not higher than at the scale of individual wetlands, despite the larger average distance among pools. Within valleys, environmental heterogeneity could not be shown to be consistently higher than within wetlands and RDA-models were not able to significantly explain beta diversity. This suggests that environmental control was probably not more important as a generator of beta diversity at this spatial scale than at the within-wetland scale. Absence of spatial patterns also suggests no important dispersal limitation within valleys.

In contrast, beta diversity among valleys tended to be relatively high. RDA-analyses demonstrated strong differences among valleys, especially for species lists. Such pattern may be generated by neutral dynamics and dispersal limitation but could also indicate control of community composition by large-scale environmental gradients. RDAanalysis on species lists indeed suggested an association between species distribution patterns with a pH-gradient, but variation partitioning revealed that the importance of large scale pH variation could not be unequivocally evaluated, whereas among-valley community differentiation proved robust. Reduced exchange of propagules among pools at this scale is indeed very plausible given the morphology of the landscape (mountain ridges, lack of hydrological connections) and the distance among valleys (ranging between 4 and 28 km). Large mammals can be important vectors for the dispersal of cladoceran dormant eggs (Vanschoenwinkel et al. 2008) and in our study area llamas Lama glama are good candidates for such zoochorous dispersal. Llamas are mainly herded in the vicinity of farms, and these animals may therefore be responsible for much of the exchange of dormant eggs between pools within wetlands and among wetlands within valleys but they may be less important as dispersal vectors among valleys.

The RDA-analyses on the relative abundance data of active communities also indicated a tendency towards differentiation among valleys, but this pattern was considerably weaker than with species lists. One can indeed expect that analyses on abundance date will be less efficient in revealing patterns of species distributions because they tend to be confounded mainly by abundance patterns of dominant and subdominant species. However, analyses on abundance data may be more sensitive to responses of communities to environmental gradients because they emphasize shifts in the relative success of species that perform differently along these gradients. In our study, the abundance data were explained by similar sets of environmental variables at the largest and smallest of spatial scales (e.g. alkalinity, surface area and sludge depth). The effect of

Table 1. Partition of variation in cladoceran communities at three levels of spatial scale using partial RDA analysis on relative abundance data of active communities and species lists (active communities and dormant propagule banks combined). E: environmental model; S: spatial model component, constructed from MEM variables; V: model component representing the 4 individual valleys; WTL: model representing the 6 individual wetlands; E/(S+V): environmental model corrected for valley identity and spatial patterns within valleys; E/(S+WTL): environmental model corrected for valley identity and spatial patterns within wetlands. R^2_{adj} (%): community variation explained by model, expressed in %; DF_{model} : degrees of freedom of model; DF_{res} : residual degrees of freedom.

		Species lists					Abundance data of active communities				
	R ² _{adj} (%)	DF _{model}	DF _{res}	F	р	R^2_{adj} (%)	DF _{model}	DF _{res}	F	р	
A) Inter-valley scale WTL and VALLEY dataset, $n = \frac{1}{2}$	59										
Ε	4.1 ^a	1	57	3.49	0.001	9.9^{d}	3	55	3.12	0.001	
V	10.6	3	55	3.19	0.005	4.1	3	55	1.82	0.020	
E/V	0.5	1	54	1.28	0.191	9	3	52	2.91	0.001	
V/E	6.5	3	54	2.38	0.001	3.3	3	52	1.69	0.027	
B) Within-valley scale VALLEY dataset. $n = 29$											
E	11.9 ^b	3	25	2.26	0.001	4.9 ^e	1	27	2.43	0.031	
S	1.3	8	20	1.05	0.389	11.9	8	20	1.47	0.072	
V	8.2	3	25	1.83	0.010	2.9	3	25	1.28	0.220	
E/(S+V)	-0.5	3	14	0.97	0.526	-0.6	1	16	0.88	0.456	
S/(E+V)	1.3	8	14	1.04	0.433	12.6	8	16	1.46	0.112	
C) Within-wetland scale PLT dataset, n =35											
E	8.5 ^c	1	33	4.15	0.001	20.8 ^f	4	30	3.23	0.001	
S	-5.6	10	24	0.82	0.907	2.4	10	24	1.08	0.339	
WTL	14.5	5	29	2.15	0.001	8.7	5	29	1.65	0.024	
E/(S+WTL)	1.6	1	18	1.36	0.196	17.4	4	15	2.25	0.001	
S/(E+WTL)	2.0	10	18	1.07	0.337	2.0	10	15	1.08	0.362	

^aenvironmental model constructed from the environmental variable pH.

^benvironmental model constructed from the environmental variables chlorophyll a, depth and total phosphorus.

^cenvironmental model constructed from the environmental variable pH.

^denvironmental model constructed from the environmental variables['] alkalinity, sludge depth and surface area.

^eenvironmental model constructed from the environmental variable TN.

^fenvironmental model constructed from the environmental variables alkalinity, sludge depth.

these variables at the largest spatial scale seemed, however, mainly a reflection of their effect at the smallest scale. Indeed, the gradient lengths of these variables were only slightly larger at the inter-valley scale than at the wetland scale, and variation partitioning based on the entire dataset showed that the explanatory power of the environmental model (E) was only slightly reduced when among-valley differences were controlled for (E/V; Table 1). This indicates that large scale environmental heterogeneity was of minor importance in explaining cladoceran community variation.

Several field studies have attempted to assess the relative importance of environmental control, mass effects, neutral processes and dispersal limitation for zooplankton metacommunities. These studies were performed on a variety of water body types with different connectivity levels at a wide range of spatial scales. From these studies, environmental control seems to come out most frequently as the dominant metacommunity organizing principle. In a system of neighboring, strongly interconnected ponds, Cottenie et al. (2003) found that environmental control generated an important degree of differentiation among communities despite the homogenizing effect of water exchange via rivulets. In a set of temporary rock pools on a small scale rock shelf, Vanschoenwinkel et al. (2007) also concluded environmental control to be the most important metacommunity structuring process, together with some indications for mass effects among interconnected pools. Pandit et al. (2009) showed the degree of environmental control to depend on the degree of habitat specialization. At a much larger spatial scale, Beisner et al. (2006) mainly found evidence for environmental control in a set of connected lakes. Although abundance data showed spatial patterns, these patterns were not reflected in the presenceabsence dataset, as abundant zooplankton species were not restricted in their spatial distribution. Based on a metaanalysis of a large number of datasets of a variety of organism groups along a broad gradient of spatial scales, Cottenie (2005) concluded that passive dispersers overall tend to track well environmental heterogeneity, although the importance of dispersal limitation appears to increase at the expense of environmental control at the larger spatial scales, probably because species fail to reach habitat patches that match with their niche requirements.

Conclusions

To our knowledge, our study is the first to simultaneously study metacommunity structure of a passively dispersing animal group at three levels of spatial scale along a range of inter-patch distances of at least two orders of magnitude. The detailed analysis of resting egg banks also allowed us to complement data obtained from active community samples to establish a high quality presence-absence dataset, yielding reliable species distribution patterns that are otherwise difficult to obtain from active community sampling alone (Vandekerkhove et al. 2005a). Our study presents a case where environmental control seems to be the dominant metacommunity structuring process at the smallest spatial scale. However, an increase in spatial scale coincided with a marked increase in beta diversity that appears to be mainly generated by dispersal limitation. Our study is well in line with the results of Cottenie (2005) but illustrates that larger spatial scales need not necessarily be associated with stronger environmental gradients, which in some cases may also explain the lack of evidence for stronger environmental control at these scales. Extrapolation of spatial diversity patterns in cladoceran metacommunities to other organism groups should be done with caution. However, we believe that our results may represent general patterns that also hold for other aquatic organisms with similar life history and dispersal strategies (e.g. short generation times, production of dormant stages, passive dispersal). Candidate organism groups are large branchiopods and small invertebrate taxa like rotifers, ostracods, turbellarians, and nematodes.

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